Aquatic Invasive Species (AIS) Early Detection Monitoring Data Form

Form 3200-xxx (R 5/2015)

Instructions: Bold fields must be completed.

Location Name	**WBIC**	**County**	**Date(s)**	**AIS sign?**	Secchi (ft or m)	Conductivity (ZM ≥ 99 umhos/cm)	**Collector(s)**	**Start Time**	**End Time**	**Total Hours** (hrs x # ppl)
John		Oconto	7/13/15	Y-stop sign no official	16ft 1'	220	M. Neult R. Motiff	1:15pm	4:15pm	

STEP 1: Circle species that you looked for and review the Identification Handout.

AQUATIC PLANTS/ALGAE					**INVERTEBRATES**		Other (please specify)
European frogbit	Hydrilla	Water hyacinth	Water chestnut	Purple loosestrife	Zebra/quagga mussels	Faucet snails	
Yellow floating heart	Curly leaf pondweed	Water lettuce	**RIPARIAN PLANTS**	Yellow flag iris	Asian clam	Chinese/Banded mystery snails	
Brazilian waterweed	Fanwort	Eurasian water milfoil	Flowering rush	Japanese knotweed	New Zealand mudsnails	Rusty/red swamp crayfish	
	Parrot feather	Didymo	Phragmites	Japanese hop		Spiny/fishhook waterflea	

STEP 2: Record locations of sampling sites (in decimal degrees). Indicate whether snorkeled or why not. List AIS found and density at each site or record none. Collect a sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate ethanol.

Site*	**Latitude**	**Longitude**	**Snorkel (Y/N)**	**If no, indicate why†**	**Species name, density (1-5)‡, and live (L) or dead (D)§**	**Sample (Y/N)**	**Photo (Y/N)**	**No AIS**	**Comments**
BL1	45.33417	-88.49818	N	shallow/ mucky	BMS-2(L)	Y			
BL2	45.33560	-88.48656	N	too dense weeds	BMS-3(L); MyoSco-1(L)	Y			
MS1	45.33698	-88.48609	—	—	BMS-1(L); MyoSco-1(L)	N			
MS2	45.33940	-88.49205	—	—	MyoSco-1(L)	N			
TS1	45.33863	-88.49641	N	too dense weeds	BMS-1(L)	N			
TS2	45.33942	-88.49220	N	" "	BMS-3(L); MyoSco-1(L)	N			
TS3	45.33698	-88.48621	N	" "	BMS-3(L); MyoSco-1(L); RC?-1(L) SolDuc-1(L)	Y			
TS4	45.33548	-88.49166	N	" "	BMS-2(L); Tiang-1(L)	Y			
TS5	45.33457	-88.49567	N	" "	BMS-2(L); SolDuc-1(L)	N			

*boat landing (BL), target site (TS), meander survey (MS).

†Stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

‡Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a while bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

§Live (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments.

Lots of NWM (genetically confirmed)

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated.

Latitude	Longitude	Method*	Net ring depth (m)	Net diameter†	Ethanol‡	Samples combined (Y or N)	Date sent
45.33667	-88.49557	obl			non	Y	
45.33670	-88.49541	"			"	"	
45.33682	-88.49528	"			"	"	

STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

Latitude	Longitude	Net ring depth (m)	Net diameter†	Ethanol‡	Samples combined (Y or N)	Date sent
45.33667	-88.49557	4m		non	Y	
"	"	"		"	"	
"	"	"		"	"	

*Horizontal, oblique, or vertical.

†30 or 50 cm.

‡Non-denatured or denatured ethanol.

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: Freckmann Herbarium, Wisconsin State Herbarium, Other _____. Date of herbarium meeting _____.
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent _____.
- Dreissenids will be sent to Science Services. Date sent _____.
- Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date _____.

STEP 6: Data was entered into SWIMS on _____ by _____

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsin.gov and Amanda.Perdzock@Wisconsin.gov).

STEP 7: Data was proofed on _____ by _____

Notes: