

Instructions: **Bold** fields must be completed.

Location Name	WBIC	County	Date(s)	AIS sign?	Secchi (ft or m)	Conductivity (ZM > 99 umhos/cm)	Collector(s)	Start Time	End Time	Total Hours (hrs x # ppl)
Leigh		Oconto	7/1/15	Ship sign trailer No New Sign		1750 260	M. Nault R. Moffitt	9:30	3:00	1 hrs

STEP 1: Circle species that you looked for and review the Identification Handout.

AQUATIC PLANTS/ALGAE	Hydrilla	Water hyacinth	RIPARIAN PLANTS	INVERTEBRATES	Other (please specify)
European frogbit	Curly leaf pondweed	Water lettuce	Flowering rush	Zebra/quagga mussels	Faucet snails
Yellow floating heart	Fanwort	Eurasian water milfoil	Phragmites	Asian clam	Chinese/Banded mystery snails
Brazilian waterweed	Parrot feather	Didymo	Japanese hop	New Zealand mudsnails	Rusty/red swamp crayfish
					Spiny/fishhook waterflea

STEP 2: Record locations of sampling sites (in decimal degrees). Indicate whether snorkeled or why not. List AIS found and density at each site or record none. Collect a sample of any new AIS found. Collect five new invasive plant specimens, 20 Dreissenids, and up to 3 of each invertebrate species. Include internal and external labels with WBIC, name of lake, county, sample date, sample type (snails, spiny water flea or zebra mussel) and collector. Legibility is appreciated. If needed, preserve with adequate ethanol.

Site*	Latitude	Longitude	Snorkel (Y/N)	If no, indicate why†	Species name, density (1-5)‡, and live (L) or dead (D)§	Sample (Y/N)	Photo (Y/N)	No AIS	Comments
BL1	45.04264	88.22366	Y	-	CMS-1(L); Tang-2(L)	Y	1		
BL2	45.04969	88.22639	Y	-	CMS-1(D); BMS-1(D);	Y	N		milfoil spp.
TS1	45.04894	88.22779	Y	-	BMS-1(D); Tang-1	Y	1		
TS2	45.05080	88.23483	Y	-		Y		X	Oxyfish
MS1	45.04951	88.24467	N	-	MyrVert? → <u>DNA</u>	Y		X	
BL3	45.04768	88.23468	N	cold!	CMS-1(L); Phag-1(L); Tang-1(L)	Y			Phage may be hypod
TS3	45.04573	88.23218	N	cold!	CMS-1(D); Phrag-1(L)	N			
TS4	45.04501	88.23544	N	cold!	CMS-1(D)				
TS5	45.04500	88.22392	Y	-	CMS-1(D)				

*boat landing (BL), target site (TS), meander survey (MS).

†stained water, turbid water, blue-green bloom, chemical treatment, other (please describe).

‡Density ratings: 1-a few plants or invertebrates, 2-one or a few plant beds or colonies of invertebrates, 3-many small beds or scattered plants or colonies of invertebrates, 4-dense plant, snail, or mussel growth in a white bay or portion of the lake, or 5-dense plant, snail or mussel growth covering most shallow areas.

§Live (L) animals will contain flesh and live plants will generally be rooted. Dead (D) animals will not contain flesh and dead plants include sterile fragments.

STEP 3: Collect Waterflea Tows from the deep hole (DH). Decant water and preserve the sample. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a completed copy of this data form, and a completed copy of the Water Flea Tow Monitoring Report (3200-128) to DNR Science Services. Legibility is appreciated.

Latitude	Longitude	Method*	Net ring depth (m)	Net diameter†	Ethanol†	Samples combined (Y or N)	Date sent
45.05071	88.23991	ob1			non	N	
45.04650	88.22638	ob1			non	Y	
45.04637	88.22666	ob1			non	Y	

STEP 4: Collect vertical Veliger Tows from 3 sites; the deep hole (DH) and two other deep areas along the downwind side of the lake. Preserve with 4 parts ethanol and 1 part sample. Submit the sample, a copy of this completed data form, and a completed copy of the Mussel Veliger Tow Monitoring Report (3200-135) to DNR Science Service. Legibility is appreciated.

Latitude	Longitude	Net ring depth (m)	Net diameter†	Ethanol†	Samples combined (Y or N)	Date sent
45.05071	88.23990	4		non	N	
45.04652	88.22638	4			Y	
45.04650	88.22638	4			Y	

*Horizontal, oblique, or vertical.
†30 or 50 cm.

‡Non-denatured or denatured ethanol.

STEP 5: Coordinate voucher and sample submission and verification with regional DNR staff for all AIS records for the specific region.

- Plants will be compiled and entered into a spreadsheet to be verified and submitted to a herbarium by an in-person appointment. Please indicate which herbarium: Freckmann Herbarium, Wisconsin State Herbarium, Other _____ Date of herbarium meeting _____
- Snails will be compiled with other regional snail specimens and sent to UW La Crosse. Date sent _____
- Dreissenids will be sent to Science Services. Date sent _____
- Crayfish compiled and sent to: Craig Roesler or Scott VanEgeren. Date _____

STEP 6: Data was entered into SWIMS on _____ by _____

Once data is entered, send scans of data sheets to central office (Maureen.Ferry@Wisconsin.gov and Amanda.Perdzock@Wisconsin.gov).

STEP 7: Data was proofed on _____ by _____

Notes:

- East Basin } West
- 50 ft hole (East Basin) } West